

## Note

### High-performance liquid chromatographic determination of macrosporin, altersolanol A, alterporriol A, B and C in fermentation of *Alternaria porri* (Ellis) Ciferri

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Macrosporin (Mac 1)<sup>1</sup>, altersolanol A (As-A 2)<sup>2</sup>, alterporriol A (Ap-A 3)<sup>3</sup>, alterporriol B (Ap-B 3)<sup>4</sup> and alterporriol C (Ap-C 4)<sup>5</sup> have been found as metabolic pigments produced by *Alternaria porri* (Ellis) Ciferri, the causal fungus of black spot disease of stone-leek (Japanese name, negi). One of these pigments, As-A, inhibits elongation of the root in seeds of lettuce and stone-leek and shows antimicrobial activity against Gram-positive and -negative bacteria<sup>6</sup>. Also, Ap-A and Ap-B were found to be atropisomers of each other<sup>3</sup>. Cladofulvin<sup>7</sup> and asphodelin<sup>8</sup> have been reported as naturally occurring  $\alpha\beta'$ -bianthraquinone. The former is a metabolic product of *Cladosporium fulvum* and the latter is a component of *Aloe saponaria* HAW. Ap-C was found to be the second example of  $\alpha\beta'$ -bianthraquinone produced by fungi. Previously, we reported the high-performance liquid chromatographic (HPLC) determination of As-A, As-B and dactylariol, reduced anthraquinones produced during the fermentation period when *Alternaria porri* was cultured on Brian's medium T<sup>9</sup>.

The structures of Ap-A, -B and -C show that they are modified bianthraquinones consisting of As-A and Mac (Fig. 1). So far as the biogenesis of Ap-A, -B and -C is concerned, two pathways can be considered, namely whether As-A and Mac are first metabolized and then bonded to alterporriols, or alterporriols are first metabolized and then their C-C linkages connecting the monomeric halves are cleaved to two halves of the molecules, As-A and Mac. This paper deals with the HPLC determina-

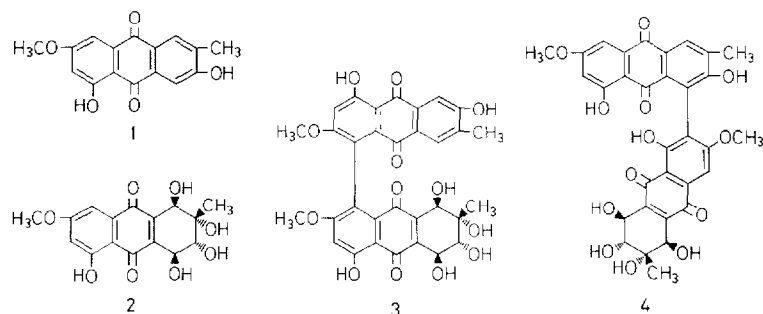


Fig. 1. Structures of pigments: 1 = macrosporin; 2 = altersolanol A; 3 = alterporriol A and B; 4 = alterporriol C.

tion of Mac, As-A and Ap-A, -B and -C during the period of fermentation in order to explore their metabolic pathways.

## EXPERIMENTAL

### *Material*

Mac, As-A and Ap-A, -B and -C were isolated as metabolic pigments of *Alternaria porri* (Ellis) Ciferri (IFO 9762), which was isolated and donated by the Institute for Fermentation, Osaka, Japan (IFO).

### *High-performance liquid chromatography*

HPLC was performed on a Shimadzu LC-6A liquid chromatograph equipped with a UV detector, operating at 254 nm for all assays. Three solvent systems were used for the resolution of five metabolic pigments, because their polarities in HPLC differed. For Ap-A, -B and -C, 0.05 M ammonium dihydrogen phosphate (adjusted to pH 2.5 with phosphoric acid) acetonitrile (7:3) was used as the mobile phase. For As-A and Mac, the same components were used but in the ratios 4:1 and 1:1, respectively. The column used was a YMC A-312 (Yamamura Chemical Laboratories), commercially packed with reversed-phase octadecylsilica (5  $\mu$ m) (150 mm  $\times$  6.0 mm I.D.), through which the above mobile phases were run at a flow-rate of 1.0 ml/min. Samples of 10  $\mu$ l were injected on to the column.

### *Fermentation and extraction of pigments*

A 2% sucrose solution of onion decoction was used as a culture medium. A number of 500-ml erlenmeyer flasks containing 200 ml of the medium were sterilized in an autoclave for 20 min at 2.3 bar and 120°C. The fungi, cultured on agar for 7–10 days, were inoculated into the flasks, which were then kept at 25°C. After fermentation for 1 day, 10 ml of the culture liquid were taken and extracted first with *n*-hexane to remove lipids and then with ethyl acetate (5  $\times$  30 ml). After evaporation of the solvent, the coloured material obtained was denoted S-1 (1.1 mg). By a similar procedure, coloured material corresponding to fermentation periods of 2, 5, 7, 14, 21 and 28 days were obtained and denoted S-2 (2.1 mg), S-3 (4.6 mg), S-4 (5.8 mg), S-5 (7.2 mg), S-6 (7.8 mg) and S-7 (8.7 mg), respectively, plus S-8 for blank (0.7 mg).

## RESULTS AND DISCUSSION

### *Quantitative analysis of Mac, As-A and Ap-A, -B and -C during the fermentation period*

As shown in Fig. 2, the retention times ( $t_R$ ) were 6.4 min (As-A,  $k' = 1.29$ ), 14 min (Mac,  $k' = 5.13$ ), 22 min (Ap-B,  $k' = 7.93$ ), 23 min (Ap-A,  $k' = 8.31$ ) and 46 min (Ap-C,  $k' = 17.61$ ). We used the internal standard method for quantitation and  $\alpha$ -naphthol ( $t_R$  26 min,  $k' = 9.89$ ) was used as the internal standard for Ap-A, -B and -C, and benzoic acid ( $t_R$  14 min,  $k' = 3.96$ ) and naphthalene ( $t_R$  19 min,  $k' = 7.53$ ) for As-A and Mac, respectively. For example, methanolic solutions of As-A (1 mg/ml) (0.4, 0.6, 0.8, 1.0 and 1.2 ml) were placed into five sample vials and 1-ml portions of methanolic solutions of benzoic acid (1 mg/ml) were added. After the volumes had been adjusted to 10 ml with methanol, 10- $\mu$ l portions of each were subjected to HPLC under the condition mentioned above. By plotting the peak-area ratios against sample weight a calibration graph for As-A was obtained. The calibration graph for Mac was



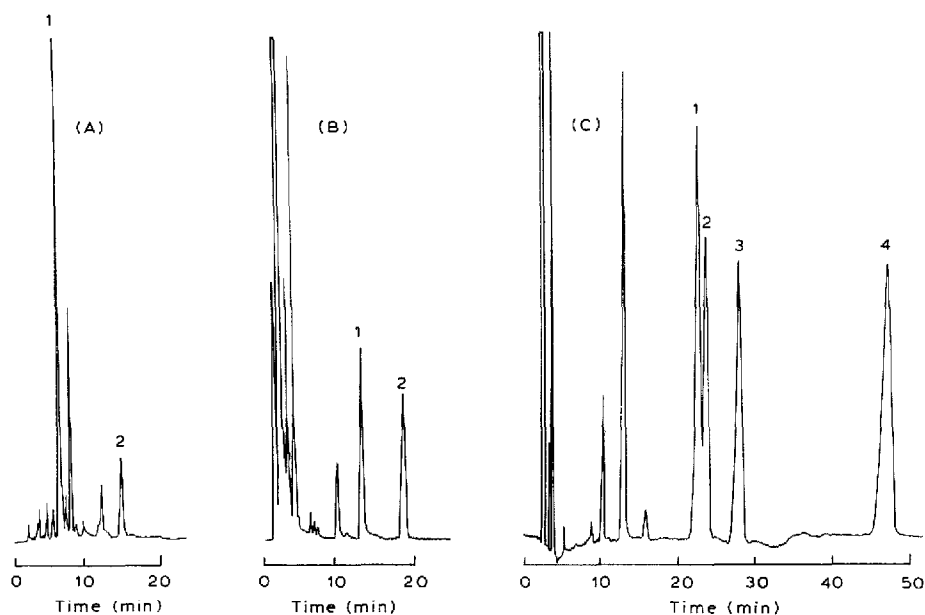


Fig. 2. Chromatograms of pigments and I.S. (A) 1 = Altersolanol A, 2 = benzoic acid; (B) 1 = macrosporin, 2 = naphthalene; (C) 1 = alterporriol B, 2 = alterporriol A, 3 =  $\alpha$ -naphthol, 4 = alterporriol C.

obtained by using the same concentration as that of As-A, whereas for Ap-A, -B and -C the concentrations were reduced 10-fold. The limits of detection, based on a signal-to-noise ratio of 10, for Mac and As-A were  $0.1 \mu\text{g/ml}$  and for Ap-A, -B and -C  $1 \mu\text{g/ml}$ .

The concentrations of Mac, As-A and Ap-A, -B and -C were calculated from the detector responses (peak areas) by using the following linear equations for the calibration graphs obtained by using the method of least squares:

$$\text{Mac: } Y = (0.589X - 2.79 \cdot 10^{-2}) \cdot 0.1$$

$$\text{As-A: } Y = (1.232X - 6.70 \cdot 10^{-3}) \cdot 0.1$$

$$\text{Ap-A: } Y = (0.748X - 2.30 \cdot 10^{-3}) \cdot 0.01$$

$$\text{Ap-B: } Y = (0.647X - 1.26 \cdot 10^{-1}) \cdot 0.01$$

$$\text{Ap-C: } Y = (0.386X + 6.24 \cdot 10^{-3}) \cdot 0.01$$

where  $Y$  is the concentration of each pigment ( $\text{mg/ml}$ ) and  $X$  is the ratio of peak area between each pigment and the internal standard (I.S.).

Extracts S-1 to S-8 were dissolved in methanol (10 ml) with the I.S. and then  $10 \mu\text{l}$  of each were subjected to HPLC under the conditions given above. The results are shown in Table I and Fig. 3. Previously, we reported that As-A was not detected in the culture liquid of the onion decoction<sup>2</sup>. In this work, however, As-A was proved to be a metabolite by means of comparison of the retention time of the metabolite with that of an authentic sample.

As shown in Table I and Fig. 3, As-A and Mac were detected after fermentation for 1 day, whereas Ap-A, -B and -C were detected after 5 days. The content of As-A in

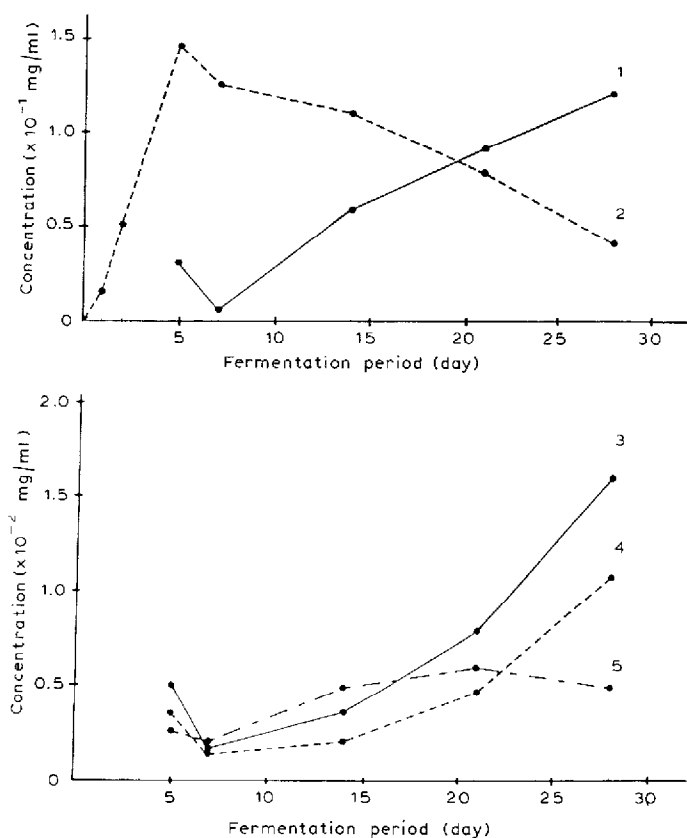


Fig. 3. Relationship between concentration of pigments and fermentation periods: 1 = macrosporin; 2 = altersolanolol A; 3 = alterporriol A; 4 = alterporriol B; 5 = alterporriol C.

the culture liquid was found to increase continuously for 7 days and then gradually to decrease, whereas Mac and Ap-A, -B and -C were found to increase gradually for up to 28 days. Previously we reported<sup>7</sup> that in the early period of fermentation As-A is first formed, and then it is converted into As-B and dactylariol when *Alternaria porri* is cultured on Brian's medium T. The present results suggest that in the early period of fermentation As-A and Mac are first formed, and then these moieties are bonded to Ap-A, -B and -C when the fungus is cultured on onion decoction medium.

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